

# TAGLN2 cDNA Cloning from *Bufo japonicus formosus* and its Diversity Analysis

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**Abstract:** During the study of bioactive polypeptides included in Bufo skin, a cDNA segment with partial transgelin-2 (TAGLN2) ORF (open reading frame) was cloned from Japanese toad Bufo japonicus formosus skin cDNA plasmid library (accession number: JX197456). To confirm the expression of the full length TAGLN2, two primers (P1 and P2) were designed based on the partial TAGLN2 sequence, and two PCR (polymerase chain reaction) reactions (P1 and XhoTT, P2 and SP6 as a pair of primers, respectively) were performed to clone the 5'- and 3'-UTR (untranslated region) using the same cDNA library as templates, and the PCR products were cloned. Based on the newly cloned 5'- and 3'-UTR sequences of TAGLN2, two new primers (P3 and P4) were designed and the second round PCR was performed by pairing P3 with XhoTT, and P4 with SP6, respectively. As the result, several TAGLN2 cDNA including full length or partial ORF were cloned indicating the expression of full length TAGLN2 in Japanese toad. Concerning its diversity, one SNP (single nucleotide polymorphism sites) in ORF was found leading to no amino acid change, and different length of 5'-UTR as well as 6 SNP in 3'-UTR was indicated.

Keywords: Bufo japonicus formosus; Transgelin-2; cDNA cloning; Diversity

## **1. INTRODUCTION**

Chan'pi (skin), Chan'yi (cortex) and Chan'su (secretions) are all *Bufo* skin-origin materials having been widely used in many prescriptions of traditional Chinese medicine and showing nice effects on pain relief, swelling and tumor control as well as many other diseases <sup>[1-4]</sup>. Cinobufacini, aqueous extract of dried *Bufo* skin, is such a clinical drug mainly aimed at the advanced cancer <sup>[5,6]</sup>. Recent studies indicated that the polypeptides purified from Cinobufacini injection showed similar antitumor activity as the injection itself in vitro experiment <sup>[7]</sup>. During the analysis of polypeptides included in *Bufo* skin, a cDNA clone with partial *transgelin-2* (*TAGLN2*) ORF was isolated from Japanese toad skin cDNA plasmid library <sup>[8]</sup>.

TAGLN2, also called SM22 $\beta$  (smooth muscle 22), is an actin binding protein which was purified from chicken gizzard smooth muscle for the first time <sup>[9,10]</sup> involving in cell proliferation and differentiation <sup>[11]</sup>, and oncogenesis <sup>[12,13]</sup>. *TAGLN2* has become a potential marker of tumorigenesis, provides a reference for early diagnosis, treatment and monitoring of tumors <sup>[14]</sup>. To confirm the expression of *TAGLN2* with full length ORF, cDNA cloning was carried out from the skin cDNA plasmid library of Japanese toad based on the partial *TAGLN2* sequence cloned previously <sup>[8]</sup>.

## 2. MATERIALS AND METHODS

## 2.1. Experimental Materials and Reagents

Japanese toad skin cDNA plasmid library held by the Japan Advanced Industrial Science and Technology (AIST, Tsukuba, Japan) was authorized Zhejiang A&F University (ZAFU) for research

as part of a Material Transfer Agreement. Concerning the library, as reported previously <sup>[15]</sup>, whose vector is pSD64TR (3250 base pairs), upstream primer is SP6 and the downstream one is S.D.A., and *Eco*R I and *Xho* I are the cloning sites. cDNA length is ranged 500-2 000 base pairs (bp). PCR kit purchased from TaKaRa; pGM-T from Tiangen (Beijing, China); the primer synthesis and DNA sequencing were done by GENEWIZ (Suzhou, China).

## 2.2. Primer Design and Cloning of TAGLN2 with Whole ORF from Japanese Toad

Based on the partial Japanese toad *TAGLN2* cDNA (GenBank accession number: JX197456)<sup>[8]</sup>, two primers (P1, P2) were designed (Table 1). P2 was paired with SP6 for the cloning of the 5'-UTR (Group 1), and P1 paired with XhoTT (a self-designed primer complementary with the area compassing the connection point of cDNA poly(A) tail and the downstream cloning site of *Xho* I) for the cloning of the 3'-UTR (Group 2) (Table 2). Based on the sequences newly cloned 5'-UTR and 3'-UTR, two other primers (P3, P4) were designed (Table 1) for the second round PCR. P4 paired with SP6 (Group 3) and P3 paired with XhoTT (Group 4) for the cloning of cDNA with original ORF (Table 2). Japanese toad skin cDNA plasmid library was used as template in the current study. All four PCR constitutions are the same except the primers as indicated in Table 2. PCR parameters are following: 94°C/3 min; (94°C/30 s, 52°C/30 s, 72°C/2.5 min) x 35 cycles. PCR products were ligated into pGM-T, and the candidates of positive clones were sequenced.

Sequences
5'-ATTTAGGTGACACTATAGAA-3'
5'-AGATCTCTCGAGTTTTTTTTTTTT-3'
5'-GCTAAAATCCAGACATC-3'
5'-GATGAGTGGATGATCTG-3'
5'-AACCACCAACCACTAAAATGG-3'
5'-TAAACATAGATTGGTTTTATT-3'

Table1. Primers used in current study

Components	Vol (µl)
10xTaq buffer	2.0
10 mM dNTPs	0.8
$Taq (5U/\mu l)$	0.2
Primer $1(2 \mu M)^*$	1.0
Primer 2(2 µM)*	1.0
Template**	0.5
$H_2O$	14.5

Table2. PCR constitutions for TAGLN2 cloning from Japanese Bufo

Notes: \* SP6 paired with P2, P1 with XhoTT, SP6 with P4 and P3 with XhoTT in Group 1, 2, 3 and 4, respectively. \*\* Japanese toad skin plasmid cDNA library

## 2.3. Homology Analysis of TAGLN2 Amino Acids

After sequencing, DNAstar/EditSeq was used to find out ORF and deduce the amino acid sequence of the encoding protein, then the amino acid sequence was applied NCBI blast program (http://blast.ncbi. nlm.nih.gov/Blast.cgi). The homolog sequences of 8 other animals were downloaded and aligned by DNAstar/MegAlign, which was also used to construct the phylogenetic tree of TAGLN2.

## 3. RESULTS

## 3.1. TAGLN2 Cloning from B. japonicus formosus

The first round PCR (lane 1 and 2 in Fig.1) was to clone *TAGLN2* cDNA with complete 5'- and 3'-UTR, and the second round PCR (lane 3 and 4 in Fig.1) was to clone the whole *TAGLN2* ORF with either complete 5'-UTR or complete 3'-UTR. Sequencing analysis indicated that 6 different clones were obtained in Group 1, 4 in Group 2, 7 in Group 3, and 6 in Group 4, totally 23 different clones were obtained. For easy description, these clones were designated as *TAGLN2*-M-N (M: Group No., N: Clone No. in each group). TAGLN2 cDNA Cloning from Bufo japonicus formosus and its Diversity Analysis



Fig1. PCR products of TAGLN2 amplified from B. japonicus formosus

M: DNA Ladder; 1: SP6 and P2; 2: P1 and XhoTT; 3: SP6 and P4; 4: P3 and XhoTT

## 3.2. TAGLN2 cDNA of B. japonicus formosus

Among the above 23 clones, *TAGLN2*-3-5 is 1 267 bp in length with 86 bp 5'-UTR, 594 bp ORF and 587 bp 3'-UTR encoding a protein consisting of 197 amino acid residues (Fig. 2), whose molecular weight is 21.953 kD, closed to that of other animal TAGLN2. Homology analysis of TAGLN2 of Japanese toad indicated 82% homology with *Xenopus laevis* (NP\_001080783.1), 81% with *Xenopus (Silurana) tropicalis* (NP\_989354.1), and 71%-78% among 6 other animals (*Rattus norvegicus*, NP\_00101314 5.1; *Crotalus adamanteus*, AFJ51813.1; *Mus musculus*, NP\_848713.1; *Homo sapiens*, AAH02616.1; *Gallus gallus*, XP\_003643901.1; *Danio rerio*, NP963870.1) (Fig. 3). So the clone was deposited into GenBank (accession number: KC820703) as *TAGLN2* of *B. japonicus formosus*. The phylogenetic tree is basically consistent with the traditional animal taxonomy (Fig. 4).

																																			1	-							→F
GA	GA	CT	TG	GA	GAG	CAG	AG	GC	TG	AT:	FCO	CAG	TC	TT	GTI	TC	CT	GC:	FGT	CT	GCI	ACT	TAC.	AAT	CAA	GA	GAC	cc	TAA	CC	AC	CAJ	ICC.	AC'	FAA	AA	TG0 M	KAJ A	AAC	AA K	AGG G	100	
TT	CA' S	TA	CG	GA( G	CTO	GAG	SCA	R	GA	AG	FC(	Q	CT	GA	AGJ K	I	GA	cci	AGA 2	AG K	TAC	CAA	ICC	CAG	E	CTO	GGA E	GA	ACA N	I	L	GG:	rgc.	AG:	rgg. W	AT.	CAA I	CA	GT(	Q	TGC	200	
CC P	TG.	AA E	TG	TG	GGG G	R	PCC F	AG	AA E	GAL	AG	AGG	GG	AA	GTI	rGG	GT	TTO	CA	GA	AG'	rgg W	CT	GAA K	GG	ACC D	GGC G	AC T	TG1	EAC	TT	AG	CA H	cc:	FAA G	TC.	AAC	TC	ccr	FTG	CCC A	300	
A.A.	TC	TG	TG	GC'	TAJ	AA2 K	I	CA	GA	CAT	rc) S	CAC	AA	TG	GCC	TI	CA	AAG	Q	AT	GGJ I	AGC	Q	GTG	TC	TC	AGT	F	CTO L	A.	GG	CC:	C	GAC	GAG R	AT	ACC	GC.	ATT	rgc A	TGC	400	
G.	AC' D	TTL	GT	TC( F	CAC	GAC 1	CAG	TT V	GA D	CT.	rg:	rgg W	GA	AG	GAJ G	K	GA D	CA:	rgg M	CT. A	AC: T	rg1 V	rcc.	AGC Q	:GG R	AC'	TCT	rga	TGA M	AC N	TT	GGG	FTG	GC: G	rtg L	GC.	AGT V	TA	CA3 T	AG K	GAC D	500	
GG	CT	GT C	TT	cce	GT( R	G G	GGG	TC	CA P	AA( N	CT(	3GI W	F	CC P	CAJ I	LAA C	AA K	TCO	CAT M	GG.	AAJ E	AAC N	AG. R	ACG	TG	CA' A	TTT F	TTC S	CCA	LAG 2	D	AAJ K	L TT	AAJ I	AAG K	AA E	GGG G	CA Q	GAG	GCG	TCA V	600	
T	CT. L	AC.	AG Q	AT M	GGG	3C2 3	T	AA N	CA	AA( K	G G	AGC	ст	s	CAG	TC	CG	GAJ G	M	AC	CGG	3C1 3	Y	GGC G	AT.	GC	CAA P	R	CAA	LAJ I	rcc	TC	rga.	ACO	GAG	CA	AAT	CA	cce	GA	AAC	700	
G.C.	AT	TA	GA	AA	AC	AGJ GCJ	LCA	TG	AT	AC'	TCA	TAT	TC	TA	AGO	TC	TT	TA	TC	TT	TC	TCC	TT	GCI	CA CT	CC	TCT GCA	TA	AAT	rG1	TA	GGG	TC SAA	CCI	TG	GT AA	CTI	TA	GC3	TAA	GAA	800 900	
AC TG	CA	AA TG	AT	CA.	AG'	TGO	AI	CG	TT	TTO	GG	CTA	AT	CT	TGO	TTA	CC	CT.	FAA	TC	CTO	GTG	AT	TGG	TG	AG	ATC	TG	CAG	TACT	IAA	GCI	IGA	GCO	JAC ACT	CT	GAA	CG	CAC	GCT	TCA	110 120	0
CT	CT	CT	GC	AT	cci	AAT	CAC	TG	TT	AA	AT	AGT	AA	TA	GTI	CI	GA	TCO	CAA	AT	AAJ	AAC	CA	ATC	TA	TG'	TTT	CA.														126	7

**Fig 2.** TAGLN2 cDNA of B. japonicus formosus with the whole ORF and the deduced amino acid sequence of the encoding protein

Underline: Start and stop codons; P1-P4: the locations of 4 primers

## 3.3. Diversity of TAGLN2 cDNA of B. japonicus formosus

As describes above, 23 clones were obtained. Due to the use of SP6 as the upstream primer, 5'-UTR sequences of the clones included in Group 1 and 3 should represent the original 5'-UTR in vivo. Meanwhile because of the use of XhoTT as the downstream primer, 3'-UTR sequences of the clones included in Group 2 and 4 should represent original 3'-UTR in vivo. Additionally, because the clones of Group 1, 3 and 4 were derived from the PCR products using SP6 paired with P2/P4, or /XhoTT paired with P3, while P2, P3 and P4, are all located in either 5'-UTR or 3'-UTR as indicated in Fig. 2, therefore, the clones from these 3 groups are supposed to have original ORF in tissue. Here, the *TAGLN2* cDNA diversity was analyzed according to ORF, 5'-UTR and 3'-UTR, respectively.

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Fig4. Phylogenetic tree of TAGLN2 between B. japonicas formosus and 8 other animals

Concerning ORF, 16 out of 19 clones from Group 1, 3 and 4 have 597 bp ORF with 7 SNP (single nucleotide polymorphism site), the remaining 3 clones have shorter ORF being the same as that reported previously <sup>[8]</sup>. Among 7 SNP, 5 appeared only once, which might be introduced by PCR. However, the remaining 2 are located at the 117<sup>th</sup> and 564<sup>th</sup> positions of ORF, which appeared repeatedly in different experiments indicating their objectivity, while they did not lead to amino acid change because of their synonymous mutations (transition between thymine and cytosine). Therefore, all 16 clones encode the same TAGLN2 of Japanese toad.

FAGLN2-3-5	GTA	GAG	ACT	TGG	AGA	CAG	AGG	CTG	ATT	CCA	GTC	TTG	TTT	CCT	GCT
FAGLN2-1-1															
TAGLN2-1-2															
FAGLN2-1-3															
FAGLN2-3-1															
FAGLN2-3-2															
FAGLN2-3-3															
FAGLN2-3-4															
TAGLN2-3-6															
FAGLN2-3-7															
FAGLN2-3-5	GTC	TGC	ACT	ACA	ATA	AGA	GAC	CCT	AAC	CAC	CAA	CCA	CTA	AAA	TGG
FAGLN2-3-5 FAGLN2-1-1	GTC	TGC	ACT	ACA	ATA	AGA	GAC	ССТ 	AAC	CAC	CAA .G.	CCA	CTA	ад <u>а</u>	TGG
FAGLN2-3-5 FAGLN2-1-1 FAGLN2-1-2	GTC	TGC 	ACT	ACA	ата 	AGA 	GAC	сст 	AAC	CAC	CAA .G.	CCA	CTA	ад <u>а</u> 	TGG
FAGLN2-3-5 FAGLN2-1-1 FAGLN2-1-2 FAGLN2-1-3	GTC	TGC 	ACT	ACA 	ата 	AGA	GAC	сст 	AAC	CAC	CAA .G. 	CCA	CTA	ада 	TGG
PAGLN2-3-5 PAGLN2-1-1 PAGLN2-1-2 PAGLN2-1-3 PAGLN2-3-1	GTC	TGC  	ACT	ACA	ATA  	AGA	GAC	сст 	AAC	CAC	CAA .G. 	CCA	CTA  	ад <u>а</u>  	<u>TG</u> G
FAGLN2-3-5 FAGLN2-1-1 FAGLN2-1-2 FAGLN2-1-3 FAGLN2-3-1 FAGLN2-3-2	GTC	TGC 	ACT	ACA  	АТА  	AGA 	GAC	сст  	AAC	CAC	CAA .G. 	CCA 	СТА 	ал <u>а</u>  	т <u>с</u> с
FAGLN2-3-5 FAGLN2-1-1 FAGLN2-1-2 FAGLN2-1-3 FAGLN2-3-1 FAGLN2-3-2 FAGLN2-3-2	GTC	TGC  	ACT	ACA	ATA  	AGA	GAC	CCT 	AAC	CAC	CAA .G. 	CCA	CTA	ад <u>а</u>  	TGG  
PAGLN2-3-5 PAGLN2-1-1 PAGLN2-1-2 PAGLN2-1-3 PAGLN2-3-1 PAGLN2-3-2 PAGLN2-3-3 PAGLN2-3-3 PAGLN2-3-4	GTC	TGC  	ACT	ACA	АТА  	AGA  	GAC	сст  	AAC	CAC	CAA .G. 	CCA	СТА 	ад <u>а</u>	TGG
TAG LN2-3-5 TAG LN2-1-1 PAG LN2-1-2 PAG LN2-1-3 PAG LN2-3-1 PAG LN2-3-2 PAG LN2-3-3 PAG LN2-3-4 PAG LN2-3-6	GTC	TGC   	ACT 	ACA 	ATA  	AGA   	GAC	сст  	AAC	CAC	CAA .G. 	CCA	СТА  	ал <u>а</u>	TGG

Fig5. Alignment of multiple TAGLN2 5'-UTR sequences of B. japonicus formosus

- : absence of corresponding nucleotide; · : identical nucleotide, Underline: Location of P3 primer;

With regard to 5'-UTR, 10 clones with full length ORF were analyzed. As shown in Fig. 5, the length is quite different among different clones, the longest one is 86 bp (*TAGLN2-3-5*), and the shortest one is only 2 bp (*TAGLN2-3-3*). But the nucleotides of the overlapped area are the same (Fig. 5). About 3'-

UTR, 11 sequences were compared and 15 SNP appeared, among which 9 appeared only once or in a certain group, most likely introduced by PCR. The remaining 6 were appeared in different experiments indicating their objective existence. Additionally, the length of poly (A) tail was different among clones (Fig. 6).

TAGLN2-2-1 TGA ACG AGC AAA TCA CCG GAA ACT CCA GAT TAC CAA ACA GAT CAT CCA CTC ATC TCC TAG GTC TTC TCT TCT TCT TGC TCA CCT TGC TCA CCT CTG [ 90] 901 901 TAGLN2-2-4 .... 901 901 901 901 901 901 901 1 901 1x197456 [180] [180] [180] [180] [180] [180] TAGLN2-4-3 TAGLN2-4-4 TAGLN2-4-5 TAGLN2-4-6 [180] [180] [180] [180] jx197456 [180] TAGLN2-2-1 TTC TTT GCA TAA TTT TTA GGG AAA CTT GAA AAA TAT TTA AAT TTT GAT TGG CTA GGA ATG AAA TGT TGT AAT GTC TTG GAG GGT ACA [270] (270) [270] TAGLN2-2-4 .... ... ... ... ... ... ... [270] TAGLN2-4-1 ... ... ... ... ... ... ... [270] [270] 12701 TACLHZ-4-3 TACLHZ-4-4 TACLHZ-4-5 TACLHZ-4-6 jx197456 [270] [270] [270] TAGLN2-2-1 CCG CAA TGC CTT TAC ATT CCA CAC CCG CCT TCC CTG TTG TCG TGT GTA TAT TTG TGA CCA AAA TCA AGT GGA TCG TTT TCT CTA ATC TTG [360] [360] [360] [360] [360] [360] 13601 (360) 13601 (360) [360] TAGLN2-2-1 GTT CCC TTA ATC CTG TGA TTG GTG AGA TCT GCA GTA AAG CAG AGC GAC CTC CAC GCA GCT TCA TGG TGA TTG AAG CAA TAG TGC AAA AGG [450] [450] [450] [450] [450] [450] [450] [450] [450] 14501 [450] TAGLN2-2-1 GAA TGC AAC GTT AAA GCT GTT CTT CGT AGT CTA TTT CCT AGT GAC TGA GGT GGG AAC TGA GAA TCC CAC GGC TCC GCT CTC TGC ATC CAA [540] TAGLN2-2-2 TAGLN2-2-3 [540] [540] TAGLN2-2-4 TAGLN2-4-1 TAGLN2-4-2 [540] [540] [540] TAGLN2-4-3 [540] TAGLN2-4-4 ... 15401 [540] [540] [540] [624] [624] [624] 16241 1x197456 [624]

Fig6. Alignment of multiple TAGLN2 3'-UTR sequences of Bufo japonicus formosus

-: absence of corresponding nucleotide; ·: identical nucleotide; Underline: Location of P4 primer; : Stop codon

#### 4. DISCUSSION

Current study clarified the expression of full length *TAGLN2* in Japanese toad skin (Fig. 2), and confirmed the existence of partial *TAGLN2* expression<sup>[8]</sup>. At the protein level, there is only one amino

acid difference between TAGLN2 of Japanese *Bufo* and or Chinese *Bufo* (*B. gargarizans*) showing homology as high as 99%. The different amino acid is located in the unimportant area, therefore, both should have the same characteristics as reported in previous study <sup>[16]</sup>. *TAGLN2* diversity of Japanese toad is mainly reflected in the length of 5'-UTR (Fig. 5), while ORF and 3'-UTR are relatively conserved (Fig. 6). Homology analysis of Japanese toad TAGLN2 showed 71-82% with other 8 vertebrates (Fig. 4), indicating its high conservancy in the process of evolution.

As an actin binding protein, TAGLN2 is involved in many physiological and pathological processes by reorganization of cytoskeleton, microfilaments<sup>[17-19]</sup>. Concerning the molecular mechanism of the antitumor activity of TAGLN2, there have been some reports that TAGLN2 noncoding region affects its posttranscriptional translation, and plays a role in inhibiting cancer metastasis together with miRNA<sup>[20-22]</sup>. But, the mechanism is still necessary to be further studied in future.

In fact, cDNA cloning either from Japanese *Bufo* or Chinese *Bufo* has been our main study to address the polypeptide ingredients working on tumor control as well as other diseases, and the expression such as *Mcl-1*, *Gal-3*, *EDF-1*, *PPDPF* and *Clu* was clarified, all of which are related with oncogenesis or metastasis, and supposed to be the anti-tumor ingredients included in *Bufo* skin-origin materials<sup>[23-27]</sup>. One more issue is that partial cDNA of *Mcl-1*<sup>[23]</sup>, *Gal-3*<sup>[24]</sup> has also been cloned. So far, several peptides derived from protein have been reported including Buforin I and II <sup>[28]</sup>, Abhisin <sup>[29]</sup> and NuBCP-9<sup>[30]</sup>, either with antimicrobial activity or antitumor activity. Combined all these studies with the report that polypeptides from Cinobufacini has antitumor activity, and the fact that most tradition Chinese medicine is taken orally, it might be not unreasonable to suggest that proteins included in tradition Chinese medicine play their pharmaceutical functions by short polypeptides. The polypeptides included in Chan'pi, Chan'yi and Chan'su, are urgent to be studied.

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